

# UK Biodiversity Indicators 2021

This document supports  
D1c. Status of pollinating Insects

## Technical background document:

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## D1c - Biodiversity and Ecosystem Services – status of pollinating insects – technical background document

Gary D. Powney, Colin A. Harrower, Louise Barwell, Charlotte Outhwaite, Nick J. B. Isaac

### Introduction

Pollination is a vital ecosystem service that benefits agricultural and horticultural production, and is essential for maintaining wild flower biodiversity. By improving the yield, quality and resilience of crops, insect pollination has been valued at £400 million per year to the UK economy (POST, 2010). Thirty five percent of the world's agricultural output, by volume, consists of 87 crop types that benefit from pollination by animals (insects, birds and mammals), but because most of these crops are not entirely dependent on animal pollination, the amount of production directly attributable to animals is lower than this value (Klein *et al.*, 2007). There is growing concern regarding the population status of insect pollinators, and in turn the pollination service they provide (Potts *et al.*, 2010; Garratt *et al.*, 2014). As with most other areas of biodiversity, the main threats to pollinators include habitat loss, environmental pollution, climate change and the spread of alien species (Klein *et al.*, 2007; Potts *et al.*, 2010; Vanbergen & The Insect Pollinators Initiative 2013). The widespread application of pesticides is also perceived as a major threat to pollinator diversity (Brittain *et al.*, 2010). In order for governments to act upon these threats they need robust metrics on the national-scale status of pollinators and pollination, though deriving such a metric has previously been limited by the availability of suitable data and analytical techniques, and the species considered to be wild pollinators are subject to debate (Hutchinson *et al.* 2021). With the increase in citizen science, the availability of large-scale biological record data has increased (Silvertown, 2009). Such data are collected without a standardized survey protocol and therefore extracting reliable trends from them can be difficult. However, with recent analytical advances it is now possible to estimate reliable trends from such data (van Strien *et al.*, 2013; Isaac *et al.*, 2014).

### Methods

#### *Data sources*

Occurrence records of bee and hoverfly species within 1km grid cells in the UK originate from the Bees, Wasps and Ants Recording Society (BWARS) and the Hoverfly Recording Scheme biological records databases. The time-period used for the indicator was 1980 to 2017, as this represents a core period of recording for these taxa in the UK. Bee species were filtered (following expert guidance from BWARS) so that only species considered to be wild pollinators were included. Species that had undergone taxonomic changes or had taxonomic issues during the time frame of the indicator were excluded from the analysis. Furthermore, models based on species with less than 50 records tend to be unreliable (Powney *et al.*, 2019), and were therefore excluded from the analysis. The final composite indicator was based on 377 species of wild pollinator, see Appendix 1 for a list of species covered. Note that the species considered to be wild pollinators are subject to review, following feedback from the scientific community and the publication of a literature review of field survey data recording wild bee visits to crops in Great Britain and Europe (Hutchinson *et al.* 2021).

#### *Generating species' occupancy models*

The data used to produce the indicator were not collected using a standardised protocol, but instead are a collation of unstructured biological observations collected by a large network of volunteer recorders. Such data tend to contain many forms of sampling bias and noise, making it hard to detect genuine signals of change (Tingley & Beissinger, 2009; Hassall & Thompson, 2010; Isaac *et al.*, 2014). Recent studies have highlighted the value of Bayesian occupancy models for estimating species occurrence in the presence of imperfect detection (van Strien *et al.*, 2013; Isaac *et al.*, 2014). This approach uses two hierarchically coupled sub-models: an occupancy sub-model (i.e. presence versus absence), and a detection sub-model (i.e. detection versus non-detection).

Together these sub-models estimate the conditional probability that a species is detected when present. Species-specific time series estimates are derived from a Bayesian occupancy model, described in Outhwaite *et al.* (2019) and following van Strien *et al.* (2013) and Isaac *et al.* (2014),

with improvements based on Outhwaite et al. (2018). Annual estimates of occupancy, with estimates of uncertainty, are available for 5,293 UK invertebrate, bryophyte and lichen species for the period 1970 to 2015 (Outhwaite et al. 2019). These models are updated as and when new data become available from recording schemes. For each site-year combination the model estimates presence or absence for the species in question given variation in detection probability: from this the proportion of occupied sites ('occupancy') was estimated for each year.

#### *Species-specific trends*

For each species, the long- and short-term trend in occupancy was estimated as the mean annual percent change (over the time-period in question) across 1,000 estimates from the posterior distribution. Species were grouped into 1 of 5 categories based on both their short-term and long-term occupancy trend (Table 1). The threshold values for each category were based on those of the wild bird indicator; whether an individual species is increasing or decreasing has been decided by its rate of annual change over the time period (long or short) of interest. If the rate of annual change would lead to an occupancy increase or decrease of between 25% and 49% over 25 years, the species is said to have shown a 'weak increase' or a 'weak decline' respectively. If the rate of annual change would lead to a population increase or decrease of 50% or more over 25 years, the species is said to have shown a 'strong increase' or a 'strong decline', respectively. These thresholds are used in the [Birds of Conservation Concern](#) status assessment for birds in the UK.

**Table 1: Thresholds used to define individual species trends**

Category	Thresholds	Threshold – equivalent
Strong increase	Above +2.81% per annum	+100% over 25 years
Weak increase	Between +1.16% and +2.81% p.a.	+33% to +100% over 25 years
Stable	Between -1.14 % and +1.16% p.a.	-25% to +33% over 25 years
Weak decrease	Between -2.73% and -1.14% p.a.	-50% to -25% over 25 years
Strong decrease	Below -2.73% p.a.	-50% over 25 years

Asymmetric percentage change thresholds are used to define these classes as they refer to proportional change, where a doubling of a species index (an increase of 100%) is counterbalanced by a halving (a decrease of 50%).

#### *Filtering out reliable occupancy estimates*

Not all model outputs can be considered reliable. A set of rules based on method exploration and testing were therefore employed to determine which index values were suitable for inclusion in a composite indicator. Species with less than 50 total records were excluded from the composite indicator, removing the rare species for which trends in distribution could not be reliability estimated. To further improve reliability, species with a gap in records greater than 10 consecutive years were excluded. In addition, to ensure a reasonable time series was available, species required at least 10 years of occupancy estimates for inclusion. Each individual species' occupancy estimates were clipped to after the first detection for the species in question.

#### *Creating a composite indicator*

Most species-based biodiversity indicators calculate the composite index as the geometric mean of indices for those species that contribute data in that year, relative to a value of 100 in the starting year.

$$I_t = 100.(\prod\{x_{1,t}, x_{2,t} \dots x_{n,t}\})^{1/n} / I_1 \quad \text{Eq 1}$$

Where  $I_t$  is the value of the indicator in year  $t$  and  $x_{1,t}$  is the index value for species 1 in year  $t$ . Under this approach, the proportional change in the indicator from one year to the next,  $\Delta_t$ , is mathematically equivalent to the geometric mean growth rate from years  $t-1$  to  $t$ .

$$\Delta_t = I_t/I_{(t-1)} = (\prod\{\lambda_{1,t}, \lambda_{2,t} \dots \lambda_{n,t}\})^{1/n} \quad \text{Eq 2}$$

$$\lambda_{i,t} = x_{i,t} / x_{i,(t-1)} \quad \text{Eq 3}$$

Where  $\lambda_{i,t}$  is the growth rate for species  $i$  from year  $t-1$  to  $t$ , and  $n$  is the number of species with index values in both year  $t-1$  and  $t$ . The geometric mean is appropriate for indices based on abundance data, which is bounded at zero but unbounded above. However, occupancy estimates are bounded at both zero and one (a species cannot occupy more than 100% of available sites). To retain this property in the indicator, the arithmetic mean of the change in log odds was selected as an appropriate statistic, thus

$$\Delta_t = I_t/I_{(t-1)} = \sum\{\gamma_{1,t}, \gamma_{2,t} \dots \gamma_{n,t}\}/n \quad \text{Eq 4}$$

$$\gamma_{i,t} = \log(p_{i,t}/(1-p_{i,t})) - \log(p_{i,(t-1)}/(1-p_{i,(t-1)})) \quad \text{Eq 5}$$

Where  $p_{i,t}$  is the proportion of occupied sites (i.e. ‘‘occupancy’’) for species  $i$  in year  $t$ , and  $\gamma_{i,t}$  is the log of the growth rate in the odds of the average site being occupied by species  $i$  between years  $t-1$  and  $t$ . Following convention, the headline indicator is set to start at 100 with a lower bound of zero:

$$I_t = 100 * \exp(\sum\{\Delta_1, \Delta_2 \dots \Delta_t\}) \quad \text{Eq 6}$$

Reformulating the composite indicator in terms of growth rates has 2 distinct advantages over the conventional approach to constructing indicators. First, it means that the categorisation of species as ‘increasing’ or ‘decreasing’ can be made from the same set of data (the growth rates) as the construction of the headline indicator. Second, it provides an elegant solution to the problem of species that join the indicator after the first year (i.e. where the first year is unreliable): other indicators typically adopt a complicated rescaling approach to ensure that species entering the indicator after the first year do not bias the overall assessment. It also makes a simple and robust, though untestable, assumption about species that drop out of the indicator prior to the final year: specifically, it assumes that their fluctuations are the same, in aggregate, as those of the species that remain in the indicator.

### Assessing the indicator line

As noted above, the index values are not classical point estimates, but rather derived from a posterior distribution of values. This distribution makes it possible to incorporate uncertainty in the annual occupancy estimates formally around the indicator line. Since each  $\gamma$  in equation 4 has a distribution of 10,000 values, it is trivial to calculate  $\Delta_t$  and  $I_t$  as a distribution, the quantiles of which measure the credible intervals of the indicator. Since this approach leads to full propagation of uncertainty from the results of each species’ model, the magnitude of uncertainty around the indicator line is expected to be larger than for other indicators, where the index values are assumed to be known without error and the uncertainty is estimated via bootstrapping. In Bayesian statistics, the magnitude of uncertainty around parameter estimates is generally referred to as the ‘credible intervals’, as opposed to the ‘confidence intervals’ derived from classical frequentist statistics. For many applications, the Bayesian credible intervals and Frequentist confidence intervals are very similar. However, the interpretation is quite different, reflecting differences in the underlying philosophy of the 2 statistical paradigms. In Frequentist statistics, the 95% confidence intervals suggest that, if an experiment were repeated many times, the true value of the parameter (e.g. the index in the most recent year) would fall within the intervals 95% of the time. In other words, the uncertainty is an expression about the data collection process, and the parameter is assumed to be fixed. By contrast, the Bayesian approach treat the data (i.e. the species’ data) as fixed and expresses uncertainty in terms of the parameter being estimated (whilst accounting for uncertainty in the data). The credible intervals around a Bayesian indicator reflect the probability that the indicator value lies within those intervals. The 90% credible intervals were chosen for making the short- and long-term assessment of a trend in the indicator line. Thus if the upper limit of the 90% credible interval falls below 100, this gives at least 95% probability (not 90%), based on the posterior distribution, of the index having declined. Thus, the long-term assessment is a simple test of whether the value 100 lies inside or outside the 90% credible intervals for the focal year. Similarly, the short-term assessment tests whether the median

value in year  $t-5$  lies within the 90% credible intervals for the focal year,  $t$ .

## Results

The indicator for all pollinators and for hoverflies has not been updated since the 2020 publication. The wild bee indicator has been extended by one additional year to 2019.

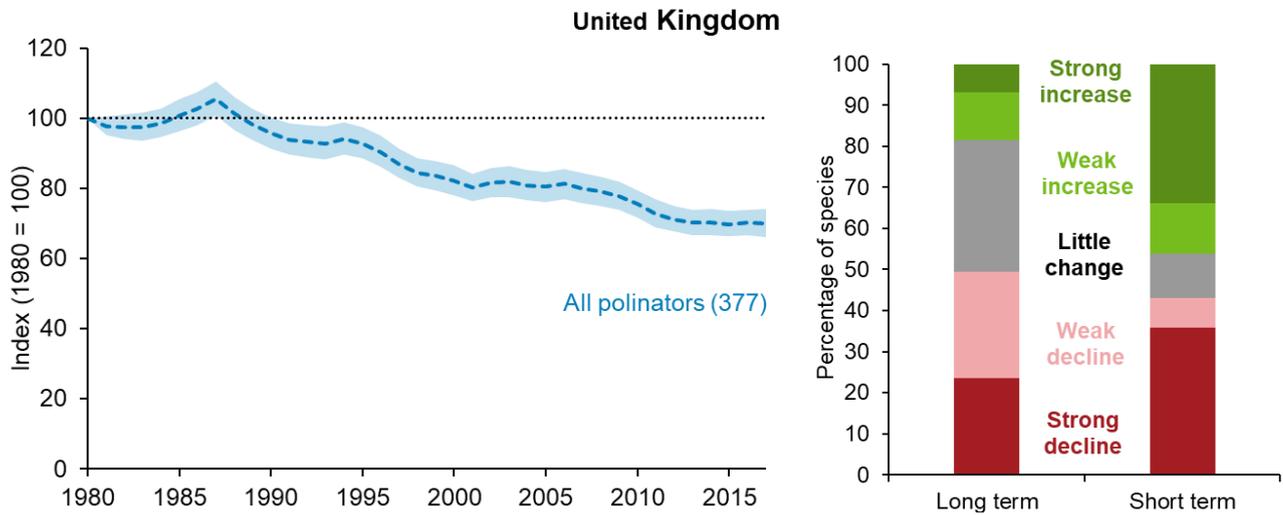
- The indicator (Figure 1) shows the average relative change in the area over which each of 377 species of pollinator was found, as measured by the number of 1km grid squares across the UK in which they were recorded – this is referred to as the ‘occupancy index’.
- Over the long term (1980 to 2017), the pollinator indicator showed 30% decline, and was therefore assessed as declining.
- Temporal patterns of change in the pollinator indicator showed a steady decline from 1987 onwards.
- Between 2012 and 2017 the indicator decreased by approximately 2%, therefore the short-term trend was assessed as little change.
- Over the long term, 19% of pollinator species became more widespread (7% showed a strong increase), and 49% became less widespread (24% showed a strong decrease).
- By contrast, over the short term, a greater proportion of species were increasing (46%; with 34% exhibiting a strong increase) than decreasing (43%; with 36% exhibiting a strong decrease).
- As individual pollinator species become more or less widespread, the communities in any given area become more or less diverse, and this may have implications for pollination as more diverse communities are, in broad terms, more effective in pollinating a wide range of crops and wild flowers.

The indicator plot was also produced for the bee (Figure 2) and hoverfly (Figure 3) species separately. Note that available data permits calculation of the indicator up to 2019 for bees, but the headline (bees and hoverflies) and hoverfly indicators only run to 2017.

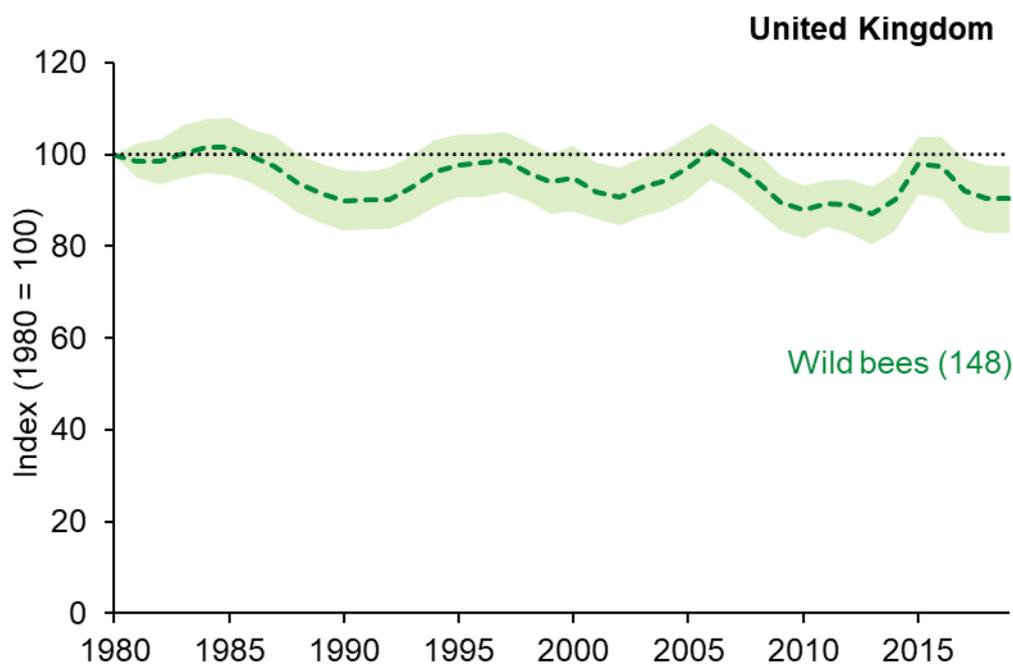
The wild bee index fluctuates around its initial value over much of the time-series. In 1983, 1985, 1997, 2006 and 2015, there is some evidence of a recovery. However, the bee index in 2019 was estimated to be 9% lower than in 1980. and is assessed as decreasing (Figure 2). A larger proportion of bee species had decreased than increased over the long term (37% decreased and 24% increased). Over the short term, the number of species decreasing (40%) and increasing (42%) is similar. The recent decline in bees is striking. A run of wet summers and agricultural practices, including pesticide use, have both been implicated in the recent decline in bees (Stanley et al., 2015). However, further research is needed to better understand the relative importance of these potential drivers of change.

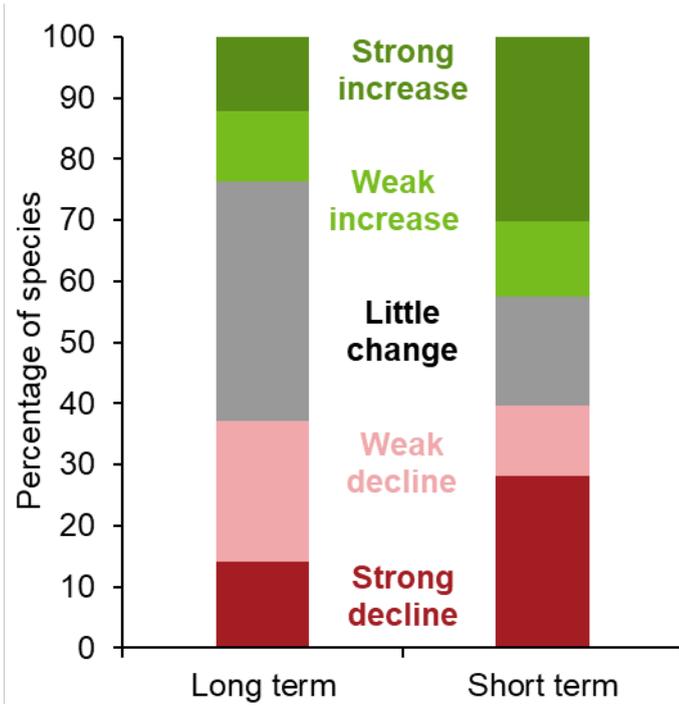
With regard to hoverflies, the index was at a peak in 1987 (109% compared to its 1980 value), and then (apart from some minor increases), underwent a progressive decline. Thus, the indicator is approximately 41% lower in 2017 than in 1980. Over the short term (2012 to 2017), the indicator decreases by just over 3%. A greater proportion of hoverflies have declined than increased in occupancy over both the long and short term (1980 to 2017: 55% decreased and 15% increased; 2012 to 2017: 49% decreased and 44% increased). It is not clear why hoverflies show a different trend to bees, although differences in the life cycle will mean they respond differently to weather events and habitat change.

**Figure 1:** Change in the distribution of wild pollinators (n = 377) in the UK between 1980 and 2017. The shaded region is the 90% credible intervals of the annual occupancy estimates and represents the uncertainty surrounding the annual estimates. The solid line illustrates the rescaled indicator value. The proportion of pollinator species in each trend category is based on the mean annual change in occupancy over both a) the long term (1980 to 2017) and b) the short term (2012 to 2017).

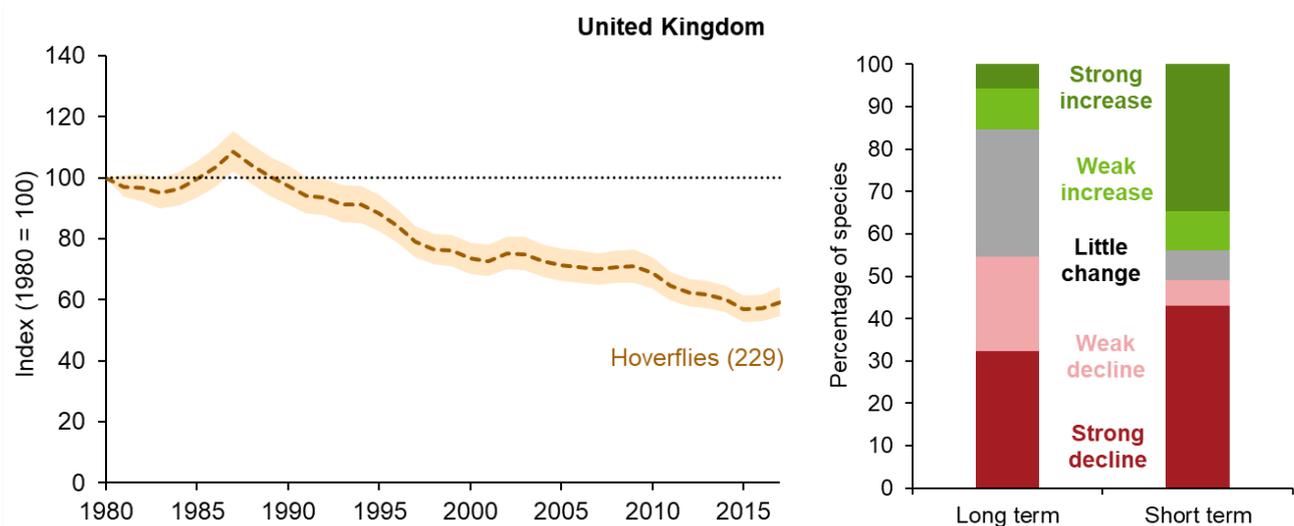


**Figure 2:** Change in the distribution of pollinating wild bee species (n = 148) in the UK between 1980 and 2019. The shaded region is the 90% credible intervals of the annual occupancy estimates and represents the uncertainty surrounding the annual estimates. The solid line illustrates the rescaled indicator value. The proportion of pollinating wild bee species in each trend category is based on the mean annual change in occupancy over both a) the long term (1980 to 2019) and b) the short term (2014 to 2019).





**Figure 3:** Change in the distribution of hoverfly species (n = 229) in the UK between 1980 and 2017. The shaded region is the 90% credible intervals of the annual occupancy estimates and represents the uncertainty surrounding the annual estimates. The solid line illustrates the rescaled indicator value. The proportion of hoverfly species in each trend category is based on the mean annual change in occupancy over both a) the long term (1980 to 2017) and b) the short term (2012 to 2017).



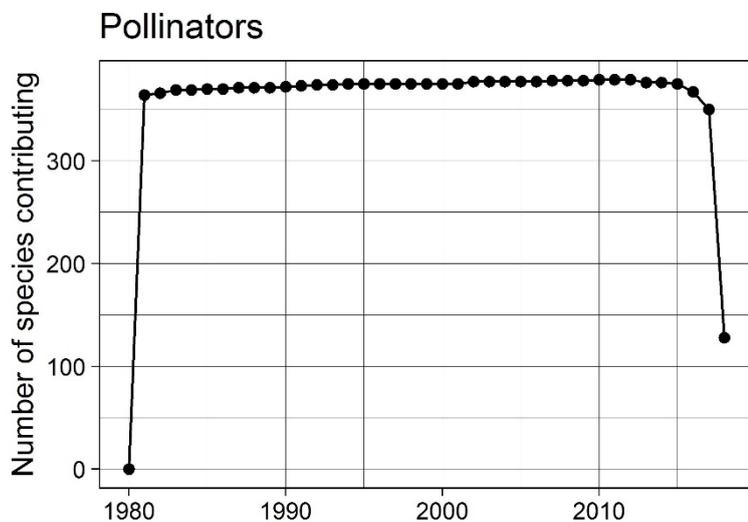
**Additional Note on The Indicator.**

Species only contribute to the indicator from the year in which they were first recorded, until the year in which they were last recorded. This means that many species only contribute for a subset of years, and hence the number of species contributing varies between years. Figure 4 shows the number of species contributing to the pollinator indicator per year up to 2018. Note that the statistical method used to produce the indicator must treat the number of species in the first year, 1980, as zero.

In 2019, the indicator spanned the period 1980 to 2016. As more data has become available for wild

bees we revisited this in 2020 to decide whether the indicator may be extended. The number of species contributing and the precision of the indicator in a given year are related; generally speaking, the more species, the better the precision. Therefore, as data becomes more sparse towards the end of the time-series (and hence fewer species contribute), we can use these measures to decide on an appropriate cut-off (final year) for the indicator. Based on the dramatic drop in number of species in 2018 – largely because there are no data for hoverflies - we do not include this year in the indicator. While there is a small drop in the number of species from 2016 to 2017 (Figure 4), the precision of the indicator is similar (Figure 1), so we decided that it is justifiable for 2017 to be the final year in the indicator time series.

**Figure 4.** Number of species contributing to the indicator in each year. Note that the statistical method used to generate the indicator requires that the start year, 1980, must be treated as having zero species.



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**Appendix : The list of the 377 species included in the pollinator indicator.****Bees**

*Andrena alfkenella*  
*Andrena angustior*  
*Andrena apicata*  
*Andrena argentata*  
*Andrena barbilabris*  
*Andrena bicolor*  
*Andrena bimaculata*  
*Andrena bucephala*  
*Andrena chrysosceles*  
*Andrena cineraria*  
*Andrena clarkella*  
*Andrena coitana*  
*Andrena congruens*  
*Andrena denticulata*  
*Andrena dorsata*  
*Andrena falsifica*  
*Andrena ferox*  
*Andrena flavipes*  
*Andrena florea*  
*Andrena fucata*  
*Andrena fulva*  
*Andrena fulvago*  
*Andrena fuscipes*  
*Andrena gravida*  
*Andrena haemorrhoea*  
*Andrena hattorfiana*  
*Andrena helvola*  
*Andrena humilis*  
*Andrena labialis*  
*Andrena labiate*  
*Andrena lapponica*  
*Andrena marginate*  
*Andrena minutula*  
*Andrena minutuloides*  
*Andrena nigriceps*  
*Andrena nigroaenea*  
*Andrena nitida*  
*Andrena nitidiuscula*  
*Andrena niveata*  
*Andrena ovatula*  
*Andrena praecox*  
*Andrena proxima*  
*Andrena rosae*  
*Andrena ruficrus*  
*Andrena scotica*  
*Andrena semilaevis*  
*Andrena similis*  
*Andrena simillima*

**Hoverflies**

*Anasimyia contracta*  
*Anasimyia interpuncta*  
*Anasimyia lineata*  
*Anasimyia lunulata*  
*Anasimyia transfuga*  
*Arctophila superbiens*  
*Brachyopa bicolor*  
*Brachyopa insensilis*  
*Brachyopa pilosa*  
*Brachyopa scutellaris*  
*Brachypalpoides lentus*  
*Brachypalpus laphriformis*  
*Caliprobola speciosa*  
*Callicera rufa*  
*Chalcosyrphus nemorum*  
*Chamaesyrphus scaevoides*  
*Cheilosia albipila*  
*Cheilosia antiqua*  
*Cheilosia barbata*  
*Cheilosia bergenstammi*  
*Cheilosia carbonaria*  
*Cheilosia chrysocoma*  
*Cheilosia cynocephala*  
*Cheilosia fraterna*  
*Cheilosia griseiventris*  
*Cheilosia grossa*  
*Cheilosia illustrata*  
*Cheilosia impressa*  
*Cheilosia latifrons*  
*Cheilosia longula*  
*Cheilosia mutabilis*  
*Cheilosia nebulosa*  
*Cheilosia nigripes*  
*Cheilosia pagana*  
*Cheilosia proxima*  
*Cheilosia pubera*  
*Cheilosia scutellata*  
*Cheilosia semifasciata*  
*Cheilosia soror*  
*Cheilosia variabilis*  
*Cheilosia velutina*  
*Cheilosia vernalis*  
*Cheilosia vicina*  
*Cheilosia vulpina*  
*Chrysogaster cemiteriorum*  
*Chrysogaster solstitialis*  
*Chrysogaster virescens*  
*Chrysotoxum arcuatum*

<i>Andrena subopaca</i>	<i>Chrysotoxum bicinctum</i>
<i>Andrena synadelpha</i>	<i>Chrysotoxum cautum</i>
<i>Andrena tarsata</i>	<i>Chrysotoxum elegans</i>
<i>Andrena thoracica</i>	<i>Chrysotoxum festivum</i>
<i>Andrena tibialis</i>	<i>Chrysotoxum verralli</i>
<i>Andrena trimmerana</i>	<i>Criorhina asilica</i>
<i>Andrena vaga</i>	<i>Criorhina berberina</i>
<i>Andrena varians</i>	<i>Criorhina floccosa</i>
<i>Andrena wilkella</i>	<i>Criorhina ranunculi</i>
<i>Anthidium manicatum</i>	<i>Dasysyrphus albostriatus</i>
<i>Anthophora bimaculata</i>	<i>Dasysyrphus friuliensis</i>
<i>Anthophora furcata</i>	<i>Dasysyrphus tricinctus</i>
<i>Anthophora plumipes</i>	<i>Dasysyrphus venustus</i>
<i>Anthophora quadrimaculata</i>	<i>Didea fasciata</i>
<i>Anthophora retusa</i>	<i>Didea intermedia</i>
<i>Bombus distinguendus</i>	<i>Doros profuges</i>
<i>Bombus hortorum</i>	<i>Epistrophe diaphana</i>
<i>Bombus humilis</i>	<i>Epistrophe eligans</i>
<i>Bombus hypnorum</i>	<i>Epistrophe grossulariae</i>
<i>Bombus jonellus</i>	<i>Epistrophe nitidicollis</i>
<i>Bombus lapidarius</i>	<i>Episyrphus balteatus</i>
<i>Bombus muscorum</i>	<i>Eriozona syrphoides</i>
<i>Bombus pascuorum</i>	<i>Eristalinus aeneus</i>
<i>Bombus pratorum</i>	<i>Eristalinus sepulchralis</i>
<i>Bombus ruderarius</i>	<i>Eristalis abusivus</i>
<i>Bombus ruderatus</i>	<i>Eristalis arbustorum</i>
<i>Bombus soroeensis</i>	<i>Eristalis horticola</i>
<i>Bombus sylvarum</i>	<i>Eristalis intricarius</i>
<i>Bombus terrestris</i>	<i>Eristalis pertinax</i>
<i>Ceratina cyanea</i>	<i>Eristalis rupium</i>
<i>Chelostoma campanularum</i>	<i>Eristalis tenax</i>
<i>Chelostoma florissomne</i>	<i>Eumerus funeralis</i>
<i>Colletes cunicularius</i>	<i>Eumerus ornatus</i>
<i>Colletes daviesanus</i>	<i>Eumerus sabulonum</i>
<i>Colletes floralis</i>	<i>Eumerus strigatus</i>
<i>Colletes fodiens</i>	<i>Eupeodes corollae</i>
<i>Colletes halophilus</i>	<i>Eupeodes latifasciatus</i>
<i>Colletes hederæ</i>	<i>Eupeodes luniger</i>
<i>Colletes marginatus</i>	<i>Eupeodes nielseni</i>
<i>Colletes similis</i>	<i>Eupeodes nitens</i>
<i>Colletes succinctus</i>	<i>Ferdinandea cuprea</i>
<i>Dasypoda hirtipes</i>	<i>Ferdinandea ruficornis</i>
<i>Eucera longicornis</i>	<i>Helophilus hybridus</i>
<i>Halictus confusus</i>	<i>Helophilus pendulus</i>
<i>Halictus rubicundus</i>	<i>Helophilus trivittatus</i>
<i>Halictus tumulorum</i>	<i>Heringia heringi</i>
<i>Heriades truncorum</i>	<i>Lejogaster metallina</i>
<i>Hoplitis claviventris</i>	<i>Lejogaster tarsata</i>
<i>Lasioglossum albipes</i>	<i>Lejops vittatus</i>
<i>Lasioglossum angusticeps</i>	<i>Leucozona glauca</i>
<i>Lasioglossum brevicorne</i>	<i>Leucozona laternaria</i>

Lasioglossum calceatum	Leucozona lucorum
Lasioglossum cupromicans	Mallota cimbiciformis
Lasioglossum fratellum	Melangyna arctica
Lasioglossum fulvicorne	Melangyna cincta
Lasioglossum laevigatum	Melangyna compositarum
asioglossum laticeps	Melangyna labiatarum
Lasioglossum lativentre	Melangyna lasiophthalma
Lasioglossum leucopus	Melangyna quadrimaculata
Lasioglossum leucozonium	Melangyna umbellatarum
Lasioglossum malachurum	Melanogaster hirtella
Lasioglossum minutissimum	Melanostoma dubium
Lasioglossum morio	Melanostoma mellinum
Lasioglossum nitidiusculum	Melanostoma scalare
Lasioglossum parvulum	Meliscaeva auricollis
Lasioglossum pauperatum	Meliscaeva cinctella
Lasioglossum pauxillum	Merodon equestris
Lasioglossum prasinum	Microdon analis
Lasioglossum punctatissimum	Microdon devius
Lasioglossum puncticolle	Myathropa florea
Lasioglossum quadrinotatum	Myolepta dubia
Lasioglossum rufitarse	Neoascia geniculata
Lasioglossum semilucens	Neoascia interrupta
Lasioglossum smeathmanellum	Neoascia obliqua
Lasioglossum villosulum	Neoascia podagrica
Lasioglossum xanthopus	Neoascia tenur
Lasioglossum zonulum	Orhonevra brevicornis
Macropis europaea	Orhonevra geniculata
Megachile centuncularis	Orhonevra nobilis
Megachile circumcincta	Paragus haemorrhous
Megachile leachella	Parasyrphus annulatus
Megachile ligniseca	Parasyrphus lineola
Megachile maritima	Parasyrphus malinellus
Megachile versicolor	Parasyrphus nigritarsis
Megachile willughbiella	Parasyrphus punctulatus
Melitta haemorrhoidalis	Parasyrphus vittiger
Melitta leporine	Parhelophilus consimilis
Melitta tricincta	Parhelophilus frutetorum
Osmia aurulenta	Parhelophilus versicolor
Osmia bicolor	Pelecocera tricincta
Osmia bicornis	Pipiza austriaca
Osmia caerulescens	Pipiza bimaculata
Osmia leaiana	Pipiza fenestrata
Osmia parietina	Pipiza lugubris
Osmia pilicornis	Pipiza luteitarsis
Osmia spinulosa	Pipiza noctiluca
Osmia uncinata	Pipizella viduata
Osmia xanthomelana	Pipizella virens
Panurgus banksianus	Platycheirus albimanus
Panurgus calcaratus	Platycheirus ambiguus
elangyna compositarum/labiatarum	Platycheirus angustatus
Melanogaster aerea	Platycheirus discimanus

Meligramma euchromum  
 Meligramma guttatum  
 Meligramma trianguliferum  
 Platycheirus clypeatus  
 Platycheirus occultus  
 Platycheirus peltatus  
 Platycheirus scutatus  
 Platycheirus splendidus  
 Sphaerophoria interrupta  
 Sphegina sibirica  
 Cheilosia caerulescens  
 Platycheirus fulviventris  
 Platycheirus granditarsus  
 Platycheirus immarginatus  
 Platycheirus manicatus  
 Platycheirus perpallidus  
 Platycheirus podagratus  
 Platycheirus rosarum  
 Platycheirus scambus  
 Platycheirus sticticus  
 Platycheirus tarsalis  
 Pocota personata  
 Portevinia maculata  
 Psilota anthracina  
 Rhingia campestris  
 Rhingia rostrata  
 Riponnensia splendens  
 Scaeva pyrastris  
 Scaeva selenitica  
 Sericomyia lappona  
 Sericomyia silentis  
 Sphaerophoria batava  
 Sphaerophoria fatarum  
 Sphaerophoria philanthus  
 Sphaerophoria rueppellii  
 Sphaerophoria scripta  
 Sphaerophoria taeniata  
 Sphaerophoria virgata  
 Sphegina clunipes  
 Sphegina elegans  
 Sphegina verecunda  
 Syrphus pipiens  
 Syrphus ribesii  
 Syrphus torvus  
 Syrphus vitripennis  
 Trichopsomyia flavitarsis  
 Triglyphus primus  
 Tropidia scita  
 Volucella bombylans  
 Volucella inanis  
 Volucella inflata  
 Volucella pellucens  
 Volucella zonaria  
 Xanthandrus comtus  
 Xanthogramma citrofasciatum  
 Xanthogramma pedissequum  
 Xylota abiens  
 Xylota florum  
 Xylota jakutorum  
 Xylota segnis  
 Xylota sylvarum  
 Xylota tarda

Xylota xanthocnema  
Baccha elongata  
Neoascia meticulosa  
Chalcosyrphus eunotus  
Cheilosia urbana  
Eristalis nemorum  
Callicera aurata  
Cheilosia albitarsis  
Cheilosia lasiopa  
Dasysyrphus pinastri  
Epistrophe melanostoma  
Eriozona erratica  
Heringia latitarsis  
Heringia pubescens  
Heringia vitripennis